

Analysis on Some Essential and Non-essential Metals in Bulla of Enset (*E. Ventricosum*) with Supporting Soil sample in West Showa Zone

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Abstract

The aim of this study was to investigate the level of essential and non essential metals in Bulla of Ensete ventricosum and soil samples in West Showa Zone. K and Na were analyzed using FAES, Ca and Mg were determined complecometric titration method and the other metals with FAAS after appropriate quality control measures were undertaken to verify and maintain the quality of the data generated. The optimized wet digestion method for Bulla and soil analysis was found effective for all of the minerals and as it was evaluated through the recovery experiment, a good percentage recovery 96.85% (Fe in Bulla) to 112% (Pb in soil) was obtained for the minerals identified. The results of this study showed that the average concentrations were ranged in order decreasing in (mgkg^{-1}) $3366.28(\text{K}) > 1881.57(\text{Na}) > 593.44(\text{Mg}) > 862.85(\text{Ca}) > 55.68(\text{Fe}) > 6.99(\text{Zn}) > 3.09(\text{Cu}) > 2.96(\text{Mn})$. Cd and Pb were not detected for Bulla and $3521.11(\text{K}) > 3497.85(\text{Mg}) > 3461.59(\text{Fe}) > 3294.93(\text{Ca}) > 1096.89(\text{Na}) > (93.99(\text{Mn}) > 16.74(\text{Zn}) > 3.77(\text{Cu}) > 0.26(\text{Cd})$ for soil sample and Pb was not detected. The concentrations of the metals were also compared with recommended maximum permissible limits and some international reports; and found to be in a good agreement indicating no exposure risk of using the bulla and corm of Ensete ventricosum under the current situation. Statistical test of significance using ANOVA revealed that there were significant differences ($P < 0.05$) between the values of metals in the bulla and soil samples obtained from all the sampling sites except Zn and Fe for bulla and Ca in soil.

Keywords: Bulla of *E. ventricosum*, Essential and non -essential metals, Soil, wet digestion

1. INTRODUCTION

Ethiopia mainly depends on agriculture. Over 85 percent of its population lives in the rural area where crop production and animal husbandry is their main stay. Enset based farming systems play an important role in food security in Ethiopia (Spring *et al.*, 1996). *Ensete ventricosum* is one of the indigenous root crops widely cultivated in the Central, South and South Western parts of Ethiopia, but recurrent droughts have led to the

expansion of Enset cultivation to other parts of the country (Brandt *et al.*, 1997; Genet *et al.*, 2004).

The genus *Ensete* comprises of 5-7 species, half of which are African, the other half Asian in origin. It is a perennial herbaceous root crop with long broad leaves and bulky pseudostem which grows wild in a number of countries in Central and Eastern Africa including Congo, Mozambique, Uganda, Tanzania and Zambia

(Genet *et al.*, 2004). *Ensete ventricosum* cultivation in Ethiopia may have started around the 15th century, south of Gibe River. Wild *Ensete ventricosum* grows in the highland altitude range (1100-3100masl) of Ethiopia in small pockets around Bonga town and the Omo River valley. In contrast, cultivated Enset grows in a wider area comprising the Central, South and South-Western parts of Ethiopia, mainly at higher altitudes (1500–3100 masl) (Shank and Chernet, 1996; Genet *et al.*, 2004). Enset is grown mainly in the South Nation Nationality Peoples Regional State followed by Oromia and Gambella Regional States (Kamar *et al.*, 2000; Almaz *et al.*, 2002).

Ensete ventricosum parts contained high percent of water (85% to 90%), which is beneficial when used as fodder during dry periods. *Ensete ventricosum* corm contained 17 of 20 amino acids. Leaves had 13% protein, among the highest available in Ethiopia, 20% crude fibre and 10% sugar. The pseudostem the main food source, was rich in 80% of soluble carbohydrates and 65% of starch, but has low protein content 4% (Mohammed *et al.*, 2013). The Bulla of Enset has low in non-essential nutrients (lead and Cadmium) and rich in essential nutrients (calcium, magnesium, potassium, zinc and iron (Minaleshewa and Chandravanshi, 2008).

The main sources of heavy metals to plant are their growth media (soil, air, nutrient solution) from which these are taken up by the roots or foliage. So plants can easily be contaminated by heavy metals in the course of cultivation or later during the processing stage. The content of heavy metals is one of the criteria for the use of plant material as food

or traditional medicines. Hence determination of mineral compositions in food and medicinal plant is essential for understanding their nutritive importance and health risk (Endashaw, 2007).

However, no literature report was found on comparative determination the concentration of essential and non-essential metals in Bulla of *Ensete ventricosum* with its supporting soil samples. Therefore, the aim of this study to compare the level of essential and non-essential metals in Bulla of *Ensete ventricosum* and soil environment from different locations of West Showa Zone, Oromia Regional State, Ethiopia by using flame atomic absorption spectroscopy and flame emission atomic spectroscopy.

2. MATERIALS AND METHODS

2.1. Instruments and Chemicals

Polyethylene plastic bags, Electronic analytical balance with 0.0001g sensitivity, A 250 ml round bottomed flasks fitted and reflux condensers. Borosilicate volumetric flasks Measuring cylinders and micropipettes, Whatman filter paper (No.42 150 mm, England), Digestive furnace (Model KDN-20C, China), Flame Photometer (ELICO, CL-378, India) and flame atomic absorption spectrophotometer FAAS (Buck Scientific Model 210 VGG, USA). The reagents and chemicals used in this study were: HNO₃ (65-68%, Uni-Chem[®] Chemical Reagent, India), HClO₄ (70-72 %, Uni-Chem[®] Chemical Reagent, India), H₂O₂ (30 %, Uni-Chem[®], India), Lanthanum chlorides hydrate (99.9%, KIRAN LIGHI laboratories, India), EDTA-Na₂ (98.5-101 %, Unic-Chem[®], India) and Stock standard of metals

2.2. Description of the Study Areas

The study areas were conducted in west Showa Zone, Oromia Regional State,

Ethiopia. It was found between 8°17'- 9°56' N and 37°01'-38°45'E and Altitude range 2000-3200 meters above sea level as indicated Table 1.

Table: 1 Geographical location and Distance from Ambo and Addis Ababa of sampling sites

Sample sites	Distance from Ambo(km)	Distance from Addis (km)	Altitude range (masl)	Latitude range (N)	Longitude range(E)
Dire Enchine	40	154	2505-2634	8°1-8°46'	37°35'-37°38'
Jeldu	72	120	2500-3200	9°02'47"-9°15'	38°05'-38°12'16"
Jibat	70	184	2000-3000	8°35'-8°50'	37°15'-37°35'

2.3.Sample Collection and Protocol

The Bulla of *Ensete ventricosum* samples were collected in January, 2016 from the three agricultural areas (woredas) with its supporting soil. Each sample was collected purposely from four different sub-sites (farm lands) to provide replicate samples. From the three agricultural areas Bulla can be prepared according to the traditional method. During harvest leaf and old leaf sheaths first removed from the edible designated plants, the edible designated plant out of the land were cut into three parts for the separation of pseudostem and Corm with knife. The leaf sheath was decorticated using a local made bamboo scraper while the leaf sheath was held on an incline at 45 to 80 degrees from the ground against a wooden plank. Then, the pseudostem was decorticated using bamboo scraper washed with distilled water and to get Bulla by squeezing pseudostem. The soil samples were

collected from the surface 15cm- 25cm (Csuros and Csuros, 2002) depth of the same four sampling areas of Enset by spade. Finally three Bulla and soil samples one from each stated areas were collected and put in clean cooled polyethylene plastic bags labeled and brought to the Ambo University laboratory.

2.4.Sample Preparation

Bulla and soil samples were collected from each sub-sites were air dried for 72hr to remove moisture and all clods and clumps. The four sub-samples were mixed equal proportion together to form a composite sample that represents each sampling areas. The powdered two samples were placed in pre-cleaned screw capped polyethylene container and stored in desiccators containing calcium chloride to keep to constant dry weight till digestion (Mitra, 2003).

using FAAS and FAES a mixture of NHO_3 and HCO_4 were used for optimization of digestion procedure for Bulla sample where as H_2O_2 in the case of soil sample as it is a strong oxidizing agent. After the optimized condition was obtained the sample were digested by using wet digestion principles.

2.5.Optimization procedure for Bulla and Soil Samples

The basic requirements for sample preparation for analysis are to get an optimum condition for digestion. To prepare a clear colorless bulla and soil sample solutions that were suitable for the analysis

2.6. Method Validation and Quality Control

2.6.1. Method Validation

2.6.1.1. Precision and Accuracy

Precision and accuracy of the analytical method was assessed by repeatability and recovery studies of matrix spike (MS). Recovery study was performed by spiking three replicate of Bulla and soil samples with a known concentration of metal standard solution (mid-range calibration concentration). The spiked samples were then subjected to the same digestion procedure like the actual sample. Precision was expressed as relative standard deviation (RSD) of the three replicate results and calculated as:

$$RSD\% = \frac{S}{\bar{X}} \times 100$$

Where RSD is relative standard deviation, S is standard deviation and \bar{X} is mean of measurements.

Accuracy is expressed as matrix spike recovery and the percent recovery results were calculated by the following equation (Javed et al., 2010).

$$\%R = \frac{C_{\text{Spike sample}} - C_{\text{Unspike sample}}}{C_{\text{added}}}$$

2.6.1.2. Instrument Detection Limit (IDL)

Instrument detection limits (IDLs) was estimated by taking seven replicate measurements of the calibration blank (Distilled water). The IDL is calculated to the concentration equal to three times the standard deviation of seven replicate measurements of blank (USEPA, 2007).

$$IDL = 3 \times S_b$$

Where, S_b is standard deviation of blank ($n=7$) and IDL is Instrument detection limit

2.6.1.3. Method Detection Limit (MDL)

Method detection Limit is the minimum concentration of analyze that can be identified measured and reported with 99% confidence that the analyze concentration is greater than zero. MDL was based up on seven replicate measurements of a series of calibration blanks (reagent blank) that are carried through the entire sample preparation scheme (Miller, 2010). The MDL was calculated by:

$$MDL = S \times T - test$$

Where, S is standard deviation of the replicated analysis with $n-1$ degree of freedom, $t = 3.71$ (T- test value for a 99% of confidence level for six degrees of freedom)

2.6.1.4. Method Quantification Limit (MQL)

Method quantification limit was obtained from analysis of seven reagents blanks which were digested in the same digestion procedure as actual samples. The method quantification limit was calculated by multiplying standard deviation of the reagent blank by ten plus the mean of the reagent blank signals (Miller, 2010)

$$MQL = \bar{X}_{\text{blank}} + 10 \times S_{\text{blank}}$$

Where, \bar{X}_{blank} is the mean of blank, S_{blank} is standard deviation of the blank.

2.6.2. Contamination Control

2.6.2.1. Method Blank

The method blank accounts for contamination that may occur during sample preparation and analysis. These could arise from the reagents, the glassware or the laboratory environment (Mitra, 2003).

Sucrose was used as matrix since there was no other plant and clear soil that can serve as the matrix for the bulla and soil samples. The blank which was prepared from the sucrose and any reagents used for the digestion was taken through the entire measurement procedure to detect contamination from reagents, sample handling, and the entire measurement process (US EPA, 2008).

2.6.2.2. Laboratory Control Sample

Laboratory Control Sample (LCS) is an aliquot of an environmental sample to which known quantities of the method analyses are added in the laboratory. The LCS would contain known concentrations of target analytes. In this study, sucrose spiked with the target analyte of known concentration the same to sample spike was carried through the same, entire sample preparation and analysis procedures used for samples. Triplicates of LCSs were analyzed along with each batch of the Bulla and soil samples. LCS recoveries for each metal of interest were calculated using (Miller, 2010; Wodaje and Alemayehu, 2014).

$$\%R = \frac{C_{found}}{C_{added}} \times 100,$$

2.7. Transfer Factor of Metals from soil to *Ensete ventricosum*

Transfer factor is the ratio of the concentration of metals in a plant to the concentration of metals in soil. Transfer factor for each metals was computed based on the method Harrison and Chirgawi (1989) described by Uwash el al., (2012) according to the following formula

$$TF = \frac{P_M \mu g g^{-1}}{S_m \mu g g^{-1}}$$

Where %R is percent recovery, C_{found} is concentration of analyte found in the sample and C_{added} Concentration of analyte added to the sample; which were used to control overall analytical method accuracy which is independent of sample matrix effects.

2.6.2.3. Matrix Spike

Matrix spike (MS) is portion of a sample spiked with known concentration(s) of target analyte(s). The spiking occurs prior to sample preparation and analysis. The purpose of a matrix spike sample is to determine whether the sample matrix contributes bias to the analytical results (Marella, 2010).

In this study, Matrix spike was prepared for each sample item by spiking aliquots of 0.5 g of each bulla, corm and soil samples with 2.5ml standards mixture solution giving concentrations of 1.0 mgL^{-1} for K, Zn, Cd and Pb; 2.0 mgL^{-1} for Na, Cu, Fe and Mn. They were all carried through the same digestion and analysis steps as an unspiked sample. And the mean recovery values of Matrix Spikes were calculated using equation.

$$\%R = \frac{C_{Spike sample} - C_{Unspike sample}}{C_{added}}$$

Where, p_m is metals concentration in plant and S_m is metals concentration in soil

2.8. Statistical Analysis

Analysis of variance (ANOVA) and F-test at $p < 0.05$ are used to examine statically significant differences in the mean concentrations of metals among groups of soil and Bulla of *E. Ventricosum*. A probability level of $p < 0.05$ is considered statistically significant. All statistical analysis was done by Microsoft Office Excel 2007 was used for data analysis and SPSS

Version 16.0 Software Window was used for Analysis of variance (ANOVA) and

3. Results and Discussion

3.1.Optimization Digestion Procedures of Soil and Bulla Samples

From the optimization procedure, digestion of 0.5 g of soil with a mixture of total volume 4.5mL which, 2ml of concentration of HNO_3 , 1.5ml concentration of HClO_4 and 1 ml concentration of H_2O_2 digested at a temperature of 230°C for 2:30 hours gave a clear colorless solution as indicated Table 2

In the same manner, the optimum conditions for bulla sample digestion were a reagent mixture of total volume 3.5mL which, 2 ml HNO_3 and 1.5 ml of HClO_4 digested at a temperature of 210°C for 2hr gave a clear colorless solution as indicated Table 3

3.2.Digestion of Bulla and Soil Samples

Applying the optimized procedures as indicated (Table 2 and 3) 0.5 g of air dried and homogenized Bulla and Soil samples were transferred into a 250 mL round bottomed flask. To this was added 3.5 ml a mixture of HNO_3 (69- 72%) and HClO_4 (70%) with a volume ratio of 2:1.5 for Bulla and 4.5 ml a mixture of HNO_3 (69- 72%), HClO_4 (70%) and H_2O (30%) with a volume As it can be show in Table 4 the mean percent recovery values ranged between 94.85 % Fe in soil to 112.8 % (Zn in Bulla), all lied in the acceptable range (80–120 %) for metal analysis (USEPA, 2008). This showed that the analytical method provided results in the required level of accuracy. The

correlation between metals in Bulla and soil samples (Miller, 2010).

ratio of 2:1.5:1 for soil. The mixture were digested on a micro Kjeldahl digestion apparatus by setting the temperature 210°C and 230°C for 2hr and 2:30 hr respectively. Then, after the digested solution were allowed to cool for 20 min without dismantling the condenser from the flask and for 10 min after removing the condenser. To the cooled solution 25 ml of distilled water was added to dissolve the precipitate formed on cooling and to minimize dissolution of filter paper by the digest residue while filtering with Whatman filter paper. The round bottom flask was rinsed subsequently with 5 ml distilled water until the total volume reached around 45 ml. To this final solution, 1% lanthanum nitrate solution was added and the solution was filled to the mark (50 ml) with distilled water. Triplicate digestions were carried out for each bulk sample. The digested samples were kept in the refrigerator, until the level of all the metals in the sample solutions were determined by FAAS and FAES.

1.1.Method Validation and Quality Control

1.1.1. Method Validation

1.1.1.1.Precision and Accuracy

RSD values of recovery was ranged between 0.05 % (Mn in corm and bulla) to 12.2 % (K in corm), all lied under the required limit ≤ 15 % (Csuros and Csuros, 2002).

Table :2 Optimization of digestion procedure for 0.5 g the Soil sample

Trial	Volume of Reagents(ml)	Time	Observation
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	Amounts of Sample	HNO ₃	HClO ₄	H ₂ O ₂	Temperature (°C)	(hr)	
1	0.5	4.0	2.0	1.0	290	3:00	Yellow
2	0.5	3.5	2	1.5	290	3:00	Light yellow
3	0.5	3.0	1.5	1.5	290	3:00	Very Clear and colorless
4	0.5	2.5	2.0	1.5	290	3:00	Light yellow
5	0.5	2.0	2.0	1.5	290	3:00	Light yellow
6	0.5	2.0	2.0	1.0	290	3:00	Very Clear and colorless
7	0.5	2.0	1.5	1.0*	290	3:00	Very Clear and colorless
8	0.5	1.5	2.0	1.0	290	3:00	Very Clear and colorless

Optimization of Temperature

1	0.5	2.0	1.5	1.0	190	3:00	Yellow
2	0.5	2.0	1.5	1.0	210	3:00	Light yellow
3	0.5	2.0	1.5	1.0	230*	3:00	Very Clear and colorless
4	0.5	2.0	1.5	1.0	250	3:00	Very Clear and colorless
5	0.5	2.0	1.5	1.0	270	3:00	Very Clear and colorless

Optimization of Time

1	0.5	2.0	1.5	1.5	230	1:45	Yellow
2	0.5	2.0	1.5	1.5	230	2:00	Light Yellow
3	0.5	2.0	1.5	1.5	230	2:15	Light Yellow
4	0.5	2.0	1.5	1.5	230	2:30*	Very Clear and colorless
5	0.5	2.0	1.5	1.5	230	2:45	Very Clear and colorless

* Optimization Condition

Table 3 Optimization of digestion procedure for the Bulla of *E. Ventricosum* sample of 0.5 g

Trial	Amounts of Sample	Volume of Reagents		Temperature (°C)	Time (hr)	Observation
		HNO ₃	HClO ₄			
1	0.5	4.0	1.5	250	2:45	Yellow
2	0.5	3.5	1.5	250	2:45	Light yellow
3	0.5	3.5	1.0	250	2:45	Very Clear and colorless
4	0.5	3.0	2.0	250	2:45	Clear and colorless
5	0.5	2.5	2.0	250	2:45	Clear and colorless
6	0.5	2.0	2.5	250	2:45	Very Clear and colorless
7	0.5	2.0	1.5*	250	2:45	Very Clear and colorless
8	0.5	1.5	1.0	250	2:45	Light yellow

Optimization of Temperature

1	0.5	2.0	1.5	150	2:45	Light yellow
2	0.5	2.0	1.5	170	2:45	Clear light yellow
3	0.5	2.0	1.5	190	2:45	Clear light yellow
4	0.5	2.0	1.5	210*	2:45	Very Clear and colorless
5	0.5	2.0	1.5	230	2:45	Very Clear and colorless

Optimization of Time

1	0.5	2.0	2.0	210	1:30	Light yellow
2	0.5	2.0	2.0	210	1:45	Clear light yellow
3	0.5	2.0	2.0	210	2:00*	Very Clear and colorless
4	0.5	2.0	2.0	210	2:30	Very Clear and colorless

* Optimization Condition

Table: 4 Recovery and precision test for the optimized procedure from sample spike (n= 3)

Metals	Sample types	Concentration in Sample (mgL ⁻¹)	Amount added (mgL ⁻¹)	Concentration in Spike (mgL ⁻¹)	Recovery (%)	Precision (% RSD)
K	Bulla	14.23±0.25	1.00	15.27±0.35	104.±10	9.62
	Soil	34.63±0.47	1.00	35.73±0.57	110±11	10.0
Na	Bulla	10.01±0.10	2.00	12.10±0.20	104.50±5	5.26
	Soil	10.43±0.057	2.00	12.63±0.115	110±2.9	2.64
Fe	Bulla	0.5830±0.015	2.00	2.52±0.196	96.85±2.3	2.30
	Soil	1.839±0.002	2.00	3.736±0.084	94.85±4.1	4.32
Mn	Bulla	0.015±0.006	2.00	2.156±0.007	107.05±0.05	0.04
	Soil	0.999±0.01	2.00	3.029±0.009	101.50±0.05	0.05
Zn	Bulla	0.077±0.006	1.00	1.205±0.009	112.80±0.30	0.30
	Soil	0.171±0.027	1.00	1.179±0.072	100.8±4.50	4.46
Cu	Bulla	0.078±0.001	2.00	2.196±0.006	105.90±0.25	0.24
	Soil	0.029±0.005	2.00	2.183±0.048	107.70±2.15	1.99
Cd	Bulla	ND	1.00	1.043±0.001	104.30±0.10	0.09
	Soil	0.004±0.001	1.00	1.009±0.005	100.50±0.40	0.04
Pb	Bulla	ND	1.00	1.062±0.007	106.20±0.70	0.70
	Soil	0.005±0.0001	1.00	1.123±0.005	112.30±0.5	0.45

RSD* = Relative standard deviation; ND* Note Detect

1.1.1.2. Instrument Detection, Method Detection and Quantification Limits

As it can be observed in Table 5 the method detection values ranged from 0.08 mgkg⁻¹ (Cd in soil) to 2.441 mgkg⁻¹ (Mn in bulla) and the MDL values lied in range from 0.07 mgkg⁻¹ (Cd in Bulla) 6.936 mgkg⁻¹ (Fe in bulla). The results revealed the both MDL and MQL values were greater than the IDL; hence, the results of the analysis could be reliable.

1.1.2. Quality Control

1.1.2.1. Calibration

Calibration curves for the various concentrations were ranged between 0.9969 and 0.9999, which were all greater than the required limit (0.995) for trace element analysis (EPA, 2007). This showed that there was good correlation (or relationship) between concentration and absorbance indicating good calibration of instrument.

Table 5: Instrument detection limit (IDL), method detection limit (MDL), method quantization limit (MQL) and correlation Coefficients of calibration curves for determined in Bulla and soil.

Metals	R ²	calibration equation	IDL(mgkg ⁻¹)	MDL(mgkg ⁻¹)		MQL (mgkg ⁻¹)	
				Bulla	Soil	Bulla	Soil
K	–	–	0.004	0.512	0.478	1.73	2.23
Na	–	–	0.003	0.45	0.465	1.70	1.72
Fe	0.9988	Y = 0.0399x + 0.0033	0.003	2.441	1.187	6.94	3.76
Mn	0.9966	Y = 0.0019x + 0.0066	0.005	2.264	1.21	7.23	3.78
Zn	0.9999	Y = 0.1188x + 0.0033	0.005	0.995	0.944	3.21	3.09
Cu	0.9999	Y = 0.788x + 0.0066	0.01	1.262	1.491	3.57	4.30
Cd	0.9999	Y = 0.1577x + 0.0033	0.005	0.09	0.08	0.07	0.90
Pb	0.9988	Y = 0.1577 + 0.0055	0.006	0.14	0.69	0.39	0.78

*IDL is Instrument detection limit, MDL is method detection limit and MQL is method quantization limit

1.1.2.2.Laboratory Control Samples Result

The percent recovery values of LCS measurements lied in the range 94.0 % (Na in soil) to 113.6 % (Zn in bulla) and their relative standard deviations 0.09 (Cu in soil) to 11.37 (Cd in soil), and all the values were found under standard control limits 80–120 % for LCS recovery, and ≤ 15 % for RSD (Csuros and Csuros, 2002; Ararso and Alemayehu, 2013). This showed that the method used for the study has provided the required level of accuracy and precision throughout the analytical process.

1.2.Level of Metals in Bulla of *Ensete ventricosum* and Soil Samples

The distribution and variations of The levels of ten metals (four major and six trace) were determined in bulla and soil samples the results were summarized in Table 6 and 7.

1.2.1. Analysis of Metals in Bulla Samples

The mean concentrations value of major nutrients (K, Na, Mg and Ca) in the bulla of Enset sample was studied areas ranged from 3213.33 to 3463.30, 1006.67 to 1336.67, 544.22 to 665.0 and 874.44 to 1041mg kg⁻¹ respectively. The mean concentrations of K in the bulla of Enset was highest than all metals analyzed. The lowest K, Na and Mg were observed in Jibat but, Na, Mg and Ca were the highest in Jeldu, which is below the maximum allowed concentration 1–5% for K, 0.1 to 0.4% for Mg and 0.1–1.0% for Ca in dry matter of plant FAO (2008) and consumption for adults, which is 2 g

sodium/day (WHO, 2008). While, as have compared the concentration of K, Na, Mg and Ca in bulla of this study greater than with the one reported by Minaleshewa and Chandravanshi (2008), which was ranged from 708 to 875 for K, 402 to 442 for Na 58.4 to 89.5 for Mg and 385 to 466mgkg⁻¹ for Ca respectively. The statistical F-test and ANOVA test showed significant variation ($p > 0.05$, at 95 % CI) between the tested bulla and soil samples

The mean concentrations value of six trace metals (Fe, Mn, Zn, Cu, Cd and Pb) in the bulla of Enset sample was studied areas ranged from 50.67 to 58.3, 1.57 to 4.676.17 to 7.40, 3.47 to 7.84 mg kg⁻¹ respectively. However, Cd and Pb could not detect from bulla sample. The concentration of iron was analyzed in the bulla of Enset samples were highest among micro nutrients. The lowest Fe and Zn were observed in Jibat but, Fe, Zn and Cu were the highest in Jeldu, which is below the maximum allowed concentration 50-250 mg kg⁻¹ for Fe, 20-300 for mg kg⁻¹ for Mn, 300 mg kg⁻¹ for Zn and 5-20 mg kg⁻¹ for Cu in dry matter of plant FAO (2008). While, as have compared the concentration of Fe, Mn, Zn and Cu in bulla of this study within the range with the one reported by Minaleshewa and Chandravanshi (2008), which was ranged from 36.5-59.8 for Fe, 1.0-4.98 for Mn, 16.3-22.0 for Zn and 2.0-3.5mgkg⁻¹ for Cu respectively. The statistical F-test and ANOVA test showed significant variation ($p > 0.05$, at 95 % CI) between the tested for soil sample except Fe and Zn no significant in bulla sample.

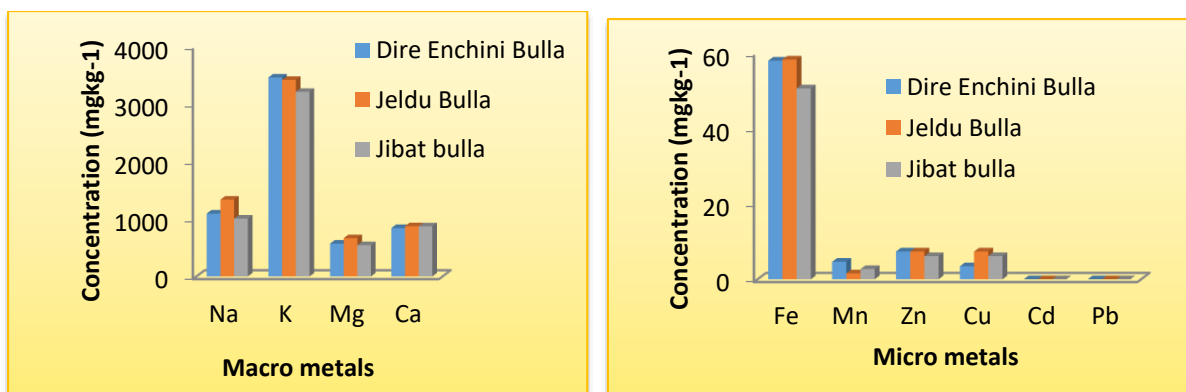


Figure: 1 Concentration of metals in Bulla of *E. Vectricosum* Sample

1.2.2. Analysis of Metals in Soil Samples

The mean Concentration of metals in soil As shown in Table 5, the trend of average concentration of metals in soil sampled from Dire Enchini is $K > Ca > Na > Mg > Fe > Mn > Zn > Cu$. Similarly, the pattern of the average concentration of metals in soil sampled from Jeldu is $Ca > K > Na > Mg > Fe > Mn > Zn > Cu$ and $Ca > K > Na > Mg > Fe > Mn > Zn > Cu$ for Jibat district. The statistical F-test and ANOVA test showed significant variation ($p > 0.05$, at 95 % CI) between the tested for soil sample except Ca. The concentrations of all metals in the

analyzed soil samples of Enset environment were under the EPA maximum permissible limit of typical concentration in soil dry matter, Fe, Mn, Cu, Zn and Cd are 10–50, 20–30, 2–100, 10–200, 0.1–1 $mg\ kg^{-1}$ respectively (EPA, 2002).

Generally, the results of all the three soil samples showed that Fe concentration was the highest and Pb was not detected. when the distribution of the selected essential and non-essential metals over each soil sample of *E. Vectricosum* was observed, they were found to vary in the order: $K > Ca > Mg > Na > Fe > Mn > Zn > Cu > Cd > Pb(ND)$ in soil samples of the three areas

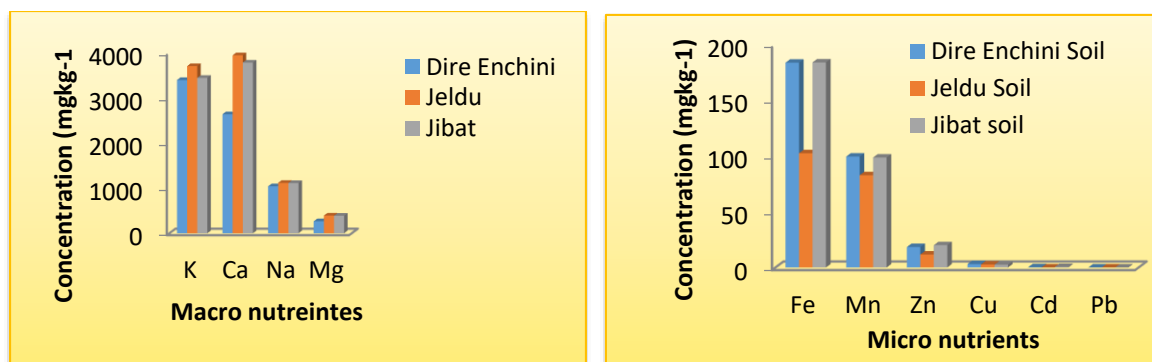


Figure: 2 Concentration of metals in Bulla of *E. Vectricosum* Sample.

Table: 6 Mean Concentration of Metals (mgkg⁻¹) in Bulla and Soil Samples from Each sites

Metals	Bulla				Soil			
	Dire Echine	Jeldu	Jibat	Optimum Range	Dire Echine	Jeldu	Jibat	Optimum Range
Na	1094.70 ±5.87	1336.67 ±647.20	1006.67 ±40.41	NA	1047.33 ±15.53	1123.3 ±20.83	1120.0 ±51.96	NA
K	3463.30 ±45.09	3422.30 ±25.17	3213.33 ±90.73	1-5% ^a	3403.33 ±5.77	3710.0 ±78.10	3450.0 ±170	NA
Mg	571.00 ±8.50	665.00 ±13.16	544.33 ±16.07	0.1-0.4% ^a	267.33 ±10.71	393.67 ±5.36	391.11 ±28.31	NA
Ca	841.67 ±14.44	874.44 ±5.00	872.44 ±8.68	0.1-1% ^a	2647.33 ±10.08	3949.1 ±43.74	3788.83 ±50.08	NA
Fe	58.03 ±0.95	58.33 ±0.58	50.67 ±4.93	50-250mgkg ^{-1a}	183.90 ±0.17	102.87 ±2.11	184.10 ±0.85	5000 ^a
Mn	4.67 ±0.58	1.50 ±0.10	2.73 ±0.21	20-300mgkg ^{-1a}	99.87 ±0.06	83.10 ±1.82	99.00 ±2.34	200 ^a
Zn	7.40 ±0.12	7.40 ±0.80	6.17 ±0.35	300mgkg ^{-1a}	18.43 ±0.67	11.67 ±0.25	20.13 ±1.21	300 ^a
Cu	3.47 ±0.50	7.84 ±0.05	4.00 ±0.01	5-20mgkg ^{-1a}	3.07 ±0.23	2.73 ±0.15	2.50 ±0.10	100 ^a
Cd	ND	ND	ND	3.0mgkg ^{-1b}	0.29±0.1	ND	0.49 ±0.01	3.00 ^a
Pb	ND	ND	ND	100mgkg ^{-1b}	ND	ND	ND	100 ^a

ND is Not Detected, NA is Not Available, ^a source: (FAO. 2008), ^b source: FAO/WHO (Codex Alimentation commission. (2001).

Table: 7 Transfer factor (TF) of Metals from Soil to *E. Ventricosum*

Sites	Sample	Metals							
		Na	K	Mg	Ca	Fe	Mn	Zn	Cu
Dire Enchini	Bulla	1.04	1.01	0.22	0.36	0.31	0.05	0.41	1.13
Jeldu	Bulla	1.18	0.92	0.22	0.22	0.56	0.018	0.6	2.8
Jibat	Bulla	0.89	0.93	0.14	0.23	0.27	0.03	0.33	1.33

1.3. Transfer Factor of Metals from soil to *E. ventricosum*

The transfer factor of metals from soil to plant in this study can be observed K, Na and Cu were more accumulated. When as compared the TF among the different metals in Bulla K, Cu and Na showed the maximum

transfer factor value in Bulla (Table 7), which ranged from 1.01 (K) in Dire Enchini) to 2.8(Cu) in Jeldu and Mn was minimum value which, ranging from 0.02 in Jeldu Bulla to 0.05 in Dire Enchini Bulla. This is indicated that Bulla is rich in essential metals.

Table 8. Comparison of the metals concentration of unprocessed Bulla of *E.ventricosum* and Soil samples were determined in this study and other literature

Metals	Uncrossed Bulla of <i>E.ventricosum</i> in this study	Processed food product Bulla of <i>E.ventricosum</i> (minaleshewa and Chandravanchi, 2008)	Metals concentration of soil in this study	Metals concentration of soil (Menegsha <i>et al.</i> , 2014)
Na	1006.7-1336.7	402-464	1073-1123	NR
K	3213.3-3463.3	708-875	3403-3710	2300.17-2382
Ca	841.3-874.4	385-446	2966.67-9633.33	55.2-56.52
Mg	544-665	58.4-89.5	469.67-543.30	914.83-939.17
Fe	50.67-58.33	36.6-59.8	102.87-184.10	NR
Mn	1.57-4.67	ND-5.38	83.10-99.87	942-953.5
Zn	6.17-7.40	2.01-3.53	11.67-20.13	55-60.97
Cu	3.47-7.84	2.01-3.53	2.50-3.03	35.46-39.67
Cd	ND	ND-5.61	NR	NR
Pb	ND	ND	ND	12.97-15

1.4. Correlation Analysis

The relationships between the essential and non essential metals were analyzed by Pearson's correlation coefficient the high correlation coefficient near +1 or -1 is a good relation between two variables and around zero no relation between them. If $r > \pm 0.7$ strongly correlation and r value between ± 0.5 to ± 0.7 moderated correlation between two different parameters (Rakesh and Raju, 2013).

As show in Table 5 strong positive correlation were observed for metals in soil between Na with Mg and Ca, K with Ca and Fe, Mg with Ca, Fe with Mg and Zn, Mn with Zn and Cd, Zn with Cd. Moderate positive correlations were observed between Na with K, K with Mg. Whereas a strong negative correlation were found between Na with Cu, K with Mn, Zn and Cd, Mg with Cu. There was also moderately negative correlation between Na with Fe and Mn, Ca with Fe and Mn). The other correlations were weak.

Table 10: Pearson's correlation coefficients between Bulla samples in three districts

	Na	K	Mg	Ca	Fe	Mn	Zn	Cd
Na	1							
K	0.59	1						
Mg	0.999*	0.55	1					
Ca	0.312	-0.583	0.358	1				
Fe	0.73	0.982	0.696	-0.421	1			
Mn	-0.599	0.293	-0.637	-0.948	0.109	1		
Zn	0.706	0.988	0.671	-0.452	0.999	0.144	1	
Cu	0.932	0.256	0.948	0.632	0.432	-0.849	0.401	1

Table 10: Pearson's correlation coefficients between soil samples in three districts

	Na	K	Mg	Ca	Fe	Mn	Zn	Cd
Na	1							
K	0.647	1						
Mg	1.00**	0.63	1					
Ca	0.997*	0.702	0.995*	1				
Fe	-0.531	0.999*	-0.512	-0.591	1			
Mn	-0.572	0.995*	-0.553	-0.631	0.999*	1		
Zn	0.363	-0.945	-0.341	-0.431	0.982	0.972	1	
Cu	0.932	-0.25	-0.91	-0.861	0.109	0.156	-0.08	1
Cd	-0.149	-0.847	-0.127	-0.271	0.915	0.894	0.974	1

*correlation is significant at the level of 0.05 levels (1-tailed) ** Correlation is significant at the level of 0.01 levels (1-tailed).

2. Conclusion

The optimized wet digestion method for Bulla and soil analysis was found effective for all of the minerals and as it was evaluated through the recovery experiment, a good percentage recovery 96% (Fe in Bulla) to 112.3% (Pb in soil) was obtained for the minerals identified. In this study the levels of ten elements in the three different district in West Showa zone. The metal contents of soil sample were found to be highest than Bulla in all analyzed except Zn, Cu and Na. The metals such as K, Ca, Mg and Na were generally found in highest concentrations than the rest metal in all of the samples.

Generally, this study showed that high level of macro elements (K, Mg, Ca and Na) were accumulated in Bulla samples collected from

the studied areas and high level of Fe from the micro nutrient followed copper. When, the distribution of the selected essential and non-essential metals over Bulla sample of *E. Vectricosum* was observed, they were found to vary in the order of decreasing K > Na > Ca > Mg > Fe > Zn > Cu > Mn in Bulla samples of the three areas. However, Cd and Pb could not be detected in Bulla. Based on the WHO recommended limit and FAO (2008) the maximum permissible limit for plant, K, Na, Mg, Ca, Fe, Mn, Cu, Zn and Pb, Cd were not found to cause any risk to the people by consuming the *E. Vectricosum* plants grown in the area where the *E. Vectricosum* is planted(studied areas). The results of all the three soil samples showed that K concentration was the highest and Pb was not detected. When, the distribution of

the selected essential and non-essential metals over each soil sample of *E. Vectricosum* was observed, they were found to vary in the order: $K > Ca > Mg > Na > Fe > Mn > Zn > Cu > Cd > Pb(ND)$ in soil samples of the three areas.

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